An Efficient Parallel Approach for Identifying Protein Families in Large-scale Metagenomic Data Sets

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Abstract-Metagenomics is the study of environmental microbial communities using state-of-the-art genomic tools. Recent advancements in high-throughput technologies have enabled the accumulation of large volumes of metagenomic data that was until a couple of years back was deemed impractical for generation. A primary bottleneck, however, is in the lack of scalable algorithms and open source software for largescale data processing. In this paper, we present the design and implementation of a novel parallel approach to identify protein families from large-scale metagenomic data. Given a set of peptide sequences we reduce the problem to one of detecting arbitrarily-sized dense subgraphs from bipartite graphs. Our approach efficiently parallelizes this task on a distributed memory machine through a combination of divide-and-conquer and combinatorial pattern matching heuristic techniques. We present performance and quality results of extensively testing our implementation on 160K randomly sampled sequences from the CAMERA environmental sequence database using 512 nodes of a BlueGene/L supercomputer.

I. INTRODUCTION

Proteins are fundamental molecules responsible for performing most of the cellular functions in an organism. Proteins that are evolutionarily- (and thereby functionally-) related are said to belong to the same "family". Identifying protein families is of fundamental importance to document the diversity of the known protein universe. It also provides a means to determine the functional roles of newly discovered protein sequences. This latter cause has become highly significant of late because numerous genome projects have been completed and as a result there is a sudden expansion of the protein universe. The most dominant contributor to this informationrevolution has been the projects in metagenomics.

Metagenomics¹ [18], the sequencing and analysis of genetic content obtained from environmental samples, is a rapidly emerging area that promises to unravel thousands of previously unknown microbial species. Among several applications, metagenomics is expected to create high impact on bioenergy, environmental biotechnology, pharmaceuticals and agriculture [17]. In a typical metagenomics project, DNA

material is collected from a target environment of interest (e.g., ocean, acid mine, human gut) and passed through a shotgun sequencing facility [22]. The collected DNA represents a pool of microbes living in that environmental sample, and the shotgun sequencing approach shreds the DNA pool into millions of tiny "fragments", each measuring only a few hundred base pairs (bp). Each of these fragments can then be individually "sequenced" in a laboratory to have its nucleotide sequence determined. The resulting environmental sequence data collection is now ready for computational analysis and discovery.

Over the last couple of years, numerous metagenomics projects have been initiated - [13], [25], [30], [31] to name a few. A continued analysis of their DNA pool is leading to collections of millions of new amino acid sequences (called ORFs for <u>Open Reading Frames</u>)² with a potential to be functional proteins. The Sorcerer II Global Ocean Sampling project (henceforth, GOS) [33], which was completed in March 2007, alone reported over 17 million new ORFs. There is an immediate need to develop software capable of processing large volumes of ORF sequences along with already known proteins (also in millions) for an accurate identification of new protein families or expansion of known families.

There are several accepted ways to define a protein family [2] — based on protein's global sequence, domain-level, or structural similarities. In this paper we consider protein families with global and domain similarities. Figure 1 illustrates an example of a protein domain family. Current approaches for detecting protein families operate by computing all-versus-all pairwise sequence similarity, and subsequently using heuristic techniques to deduce family relationships from pairwise similarities [3], [26], [33]. There are two key limitations to this approach: (i) Computing all-versus-all pairwise similarities could become computationally prohibitive even for hundreds of thousands of sequences because for n sequences the runtime complexity is $\Omega(n^2)$. This high complexity is often times offset by compromising the quality of the output; and (ii) The

¹The area has also come to be known as environmental genomics and microbial community genomics.

²Henceforth, we use the terms "ORFs" and "amino acid sequences" interchangeably.

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	10	20	30	40	50	60	7
	1			1	1	1	
d106ua3	PPEVIQQYLS	GGMCG.YDLDG	CPVWYDIIGP	LDAKGLLFSA	SKQDLLRTKM	RECELLLQEC	AHQTTKLGR
ENSP00000316203	PPEVIQQYLS	GGMCG.YDLDG	CPVWYDIIGP	LDAKGLLFSA	SKQDLLRTKM	RECELLLQEC	AHQTTKLGR
ENSP00000215812	wqPPEVIQKYMP	GGLCG.YDRDG	CPVWYDIIGP	LDPKGLLFSV	TKQDLLKTKM	RDCERILHEC	DLQTERLGK
ENSP00000255858	PPEVIQLYDS	GGLCG.YDYEG	CPVYFNIIGS	LDPKGLLLSA	SKQDMIRKRI	KVCELLLHEC	ELQTQKLGR
ENSP00000275162	IKQALKDGFP	GGLAN.LDHYG	RKILVLFAAN	WDQSRY	TLVDILRAIL	LSLEAMI	EDPEL
ENSP00000357422	IKQALKDGFP	GGLAN.LDHYG	RKILVLFAAN	WDQSRY	TLVDILRAIL	LSLEAMI	EDPEL
ENSP00000260116	SIIGLLKAGY	HGVLRsRDPTG	SKVLIYRIAH	WDPKVF	TAYDVFRVSL	ITSELIVQE-	VET
ENSP00000361995	LKDVLASGF	LTVLPhTDPRG	CHVVCIRPDR	WIPSNY	PITENIRAIY	LTLEKLIQS-	EET
ENSP00000262605	LKDVLASGF	LTVLPhTDPRG	CHVVCIRPDR	WIPSNY	PITENIRAIY	LTLEKLIQS-	EET
ENSP00000325506	IKRALIDGFP	GVLEN.RDHYG	RKILLLFAAN	WDQSRN	SFTDILRAIL	LSLEVLI	EDPEL

Fig. 1. A partial alignment of the CRAL/TRIO domain family of proteins, which contains 51 protein members (not all shown). The alignments are from the SUPERFAMILY database of structural and functional protein annotations [14].

algorithms to deduce family relationships from pairwise similarities store all pairwise results making the space complexity $\Theta(n^2)$. While such high complexity in time and space should make the problem an ideal candidate to benefit from parallel computing, there are hardly any parallel approaches. Even those that deploy parallelism resort to brute-force allocation of tasks across multiple computers and to using specialized largememory high-end platforms for tackling the space problem. For example, a recent analysis [33] of ~28.6 million ORFs took an aggregate 10^6 CPU hours. The task was parallelized using 125 dual processors systems and 128 16-processor nodes each containing between 16GB-64GB of RAM.

In this paper, we present the development of a novel parallel approach suited to exploit the aggregate memory and compute power of large scale distributed memory machines for identifying protein families in large-scale inputs. The key strengths of our approach are as follows: (i) Using pattern matching heuristics our approach overcomes the necessity to compute all-pairs sequence similarities thereby resulting in a drastic computation reduction in practice; (ii) To overcome the memory bottleneck, our approach breaks a large problem instance into numerous small sub-problems, such that each sub-problem can be solved individually on one compute node. This divide-and-conquer step is also parallel with an overall linear memory requirement in the input size; and (iii) To ensure high quality output, our approach reduces the problem to one of finding dense subgraphs in bipartite graphs wherein there exist several well-studied approximation algorithms. Our approach is built on ideas from methods developed earlier for two other applications — genome assembly [19] and dense subgraph detection for internet data [12].

In what follows, we evaluate existing approaches (Section II), outline our approach (Section III), describe our algorithms (Section IV), and present and discuss the results of our performance and quality experiments (Section V). Sections VI and VII summarize the findings and provide new directions for further research.

II. EXISTING APPROACHES

Several public repositories are available for documenting proteins along with their family and functional annotation information [2], [8], [11], [14]–[16], [28]. Protein family relationships are based on sharing conserved domains [3] or full-length similarity [26], [33]. Given two sequences, either of these relationships can be evaluated in time proportional to the product of sequence lengths using dynamic programming [23], [27] or even faster using approximate techniques such as BLAST [1]. Both these techniques account for sequence mismatches and other differences while trying to maximize matches.

To the best of our knowledge, the GOS project [33] is the only work that implements a large-scale methodology for protein family identification in metagenomic data. Given ninput ORF sequences, the GOS approach can be outlined as follows:

- Redundancy removal: Sequences that are more than 95% contained in another sequence are deemed redundant and are eliminated from further analysis. This is accomplished by performing all-versus-all sequence comparison using the BLASTP program [1].
- 2) Graph generation: A graph is created with vertices representing the set of non-redundant sequences. An edge is created between two vertices i and j if and only if sequences i and j share a "significant" sequence similarity. The graph is constructed by running the BLASTP program on all pairs of sequences using a corresponding similarity cutoff. In their implementation, the GOS team reports using a 70% similarity cutoff [33].
- 3) **Dense subgraph detection:** Because of the expectation that each sequence in a family is similar to a majority of its family members, the next step is to analyze the graph for "dense" subgraphs. However, as a related optimization problem is NP-Hard [9], the GOS approach adopts a heuristic strategy of creating sequence core sets of a bounded size, expanding each core set using a more relaxed criteria, and merging expanded sets that intersect.

For parallelization, the set of BLASTP tasks in step (1) are distributed in parallel to multiple computers. Steps (2) and (3) require creation of adjacency list and random access and therefore are implemented using large-memory SMP nodes. While this approach has been successfully applied to the



Fig. 2. Our parallel approach for protein family identification. Dotted boxes represent dense subgraphs.

largest data collection to date, the parallelization strategy can best be described as ad-hoc. Also, the graph generation and analysis in steps (2) and (3) imply $\Theta(n^2)$ storage. Furthermore, the core set creation and expansion schemes in step (3) are both such that two sequences are grouped together if they share some k neighbors. Due to computational limitations the value of k is restricted to 10. This strategy has the potential to report subgraphs that are not well connected. Ideally, k should grow with the size of the subgraph.

III. OUR APPROACH TO THE PROTEIN FAMILY IDENTIFICATION PROBLEM

The goals for our work are to (i) eliminate the $\Theta(n^2)$ space requirement, (ii) enable the use of large-scale distributed memory machines for the problem, and (iii) provide a high quality dense subgraph detection scheme that is also practically efficient. Our parallel approach, illustrated in Figure 2, consists primarily of four main phases: (i) redundancy removal, (ii) detection of connected components, (iii) bipartite graph generation, and (iv) detection of dense subgraphs.

The main idea is as follows. Given a set of n non-redundant ORF sequences, we define an undirected graph such that the vertices represent the set of n sequences and an edge is drawn between two vertices if and only if the corresponding sequences satisfy a user-specified similarity cutoff. Our approach is to devise a strategy to detect dense subgraphs without having to build or store the entire graph, so that it becomes practically possible to achieve $o(n^2)$ run-time and space complexities. This is a significant improvement over the strategy deployed by the GOS project, wherein the dominant computation and memory bottleneck is due to full graph construction and storage. Our approach is to first dynamically generate a set of connected components in parallel by using a combination of pattern matching and clustering heuristics developed for a related problem of DNA sequence clustering and genome assembly [19]. As dense subgraphs cannot span across multiple connected components, it is sufficient to subsequently distribute the connected components across multiple processors in a load balanced fashion and analyze each connected component without requiring to communicate. This allows breaking a large problem instance into multiple disjoint smaller sub-problems. While there exists a theoretical worstcase of the graph containing just one connected component, practical observations rule out such possibilities. In fact, the GOS project reported \sim 300K clusters with only as few as 542 clusters of size 2,000 or more [33].

After connected components are detected and assigned to individual processors, the next step is to identify an arbitrary set of disjoint dense subgraphs from each connected component. The disjoint aspect of the subgraphs is due to the expected many-to-one mapping from proteins to families. Let $G(V = \{s_1, s_2, \dots, s_n\}, E)$ denote a connected component. To detect dense subgraphs in G, we first transform G into a bipartite graph and then solve the dense subgraph problem for bipartite graphs. This strategy allows us to encapsulate two variations in the definition of protein family membership. In the first version, two proteins are said to be members of the same family if they share a significant global similarity to one another. In the second version, two proteins can be linked as family members if they share a substantial number of exact word matches (or domains) between them. The biological relevance of these two membership variations stems from the fact that it is possible to have both global and domain-level similarities across multiple proteins belonging to the same family [2]. Consequently, we describe two different dense subgraph detection formulations that capture these two variants of the problem. Figure 3 illustrates our approach for the two formulations.

The global similarity-based approach: Any undirected graph G(V, E) can be transformed into an equivalent undirected bipartite graph $B_d(V_l, V_r, E')$ with duplicated vertices such that $|V_l| = |V_r| = |V|$ and $E' = \{(i, j), (j, i) | (s_i, s_j) \in E\}$. Informally, the problem of finding dense subgraphs in G is therefore equivalent to finding subsets $A \subseteq V_l$ and $B \subseteq V_r$ such that A and B are well connected and $\frac{|A \cap B|}{|A \cup B|} \ge \tau$ for some cutoff $0 << \tau \le 1$. A corresponding optimization problem is NP-Hard [9]. However, a similar problem has been addressed before in the context of finding web communities and practically efficient strategies exist [10], [12]. The subtle difference is that for web communities a dense subgraph is a pair of subsets A and B such that the each vertex (or website) in B is pointed by a majority of vertices in A, without necessitating $A \approx B$. In other words, a dense subgraph in B_d as per our definition can be expected to be detected by an



Fig. 3. Illustration of the two dense bipartite subgraph reduction schemes. Dotted boxes represent dense subgraph outputs.

algorithm that solves the corresponding web community dense subgraph problem. But not every dense subgraph reported by the latter may satisfy our $A \approx B$ criterion. But this is an added constraint that can be tested in a post-processing step. This is the main idea behind our "global similarity-based" approach. We use the algorithm by Gibson *et al.* [12] for generating our shortlist of dense subgraphs because it is suited for very large inputs. Henceforth, we refer to this algorithm as the "Shingle" algorithm.

The domain-based approach: This approach makes a different bipartite graph reduction which stems from the following observation: Protein sequences that share multiple domains can also be expected to share substantially long and possibly non-contiguous exact matches (as shown in the example in Figure 1). Therefore, if multiple sequences share a sufficiently large number of fixed-length exact matches, then they are likely to belong to the same family. Thus fixed-length exact matches can be used as supporting evidence to group ORF sequences together. To implement this idea, we consider the following bipartite graph construction: Given G = (V, E) as before, let $V_m = \{e_1, e_2, \dots, e_m\}$ denote the set of all w-length strings that are present as substrings in at least two different input sequences s_i and s_j , for some fixed $w \approx 10$. Let $V_r = V$. Then construct a bipartite graph $B_m = \{V_m, V_r, E'\}$ such that $E' = \{(e_i, s_j) \mid e_i \text{ is a substring in } s_j\}$. This construction makes the dense subgraph problem equivalent to the web community version because in the latter's context websites are grouped based on the supporting evidence provided by inlinking websites. Hence, for any pair of subsets $A \subseteq V_m$ and $B \subseteq V_r$ reported by the Shingle algorithm, our desired dense subgraph output is B.

IV. Algorithms for Parallel Detection of Protein Families

Let $S = \{s_1, s_2, \dots, s_n\}$ be the set of *n* input ORFs; and $\ell = \sum_{i=1}^{n} \frac{|s_i|}{n}$. Let *p* denote the number of processors.

A. Redundancy Removal

Definition 1: Sequence s_i is said to be "contained" in s_j if an optimal alignment of s_i with s_j satisfies the following properties: (i) the similarity of the overlapping region is at least

95%; and (ii) at least 95% of s_i is included in the overlapping region³.

Problem 1: Given S, remove any s_i that is contained in any other sequence.

While this problem can be solved by comparing each pair of sequences as in the all-versus-all strategy, such an approach would take $\Theta(n^2 \ell^2)$ run-time. Instead, we apply a standard pattern matching heuristic of designing an exact-match based filtering technique for shortlisting a set of sequence pairs that show high promise to exhibit sequence similarity, and later performing alignment computations [27] only on this reduced subset. This is achieved by first constructing a generalized suffix tree (GST) [21] data structure on S and then using it as a string index for detecting sequence pairs that share a maximal match of a specified length ψ or greater. To generate pairs containing maximal matches, we use the optimal parallel algorithm described in [19]. The value of ψ is determined by the expected rates of mutation and error tolerance. For example, if two sequences aligning over a length of 100 characters must contain at least 98% similarity then they can differ in at most 2 positions, implying that there should exist at least one matching segment of length 33 characters or more. Therefore, a 33-character or longer exact match can be used as a necessary but not sufficient condition for sequence similarity. The run-time complexity of this phase is $O(\frac{n\ell}{p} + \text{#pairs generated})$, and the space complexity is $O(\frac{n\ell}{p})$. The parallel GST construction algorithm runs in $O(\frac{n\ell^2}{p})$ and requires $O(\frac{n\ell}{p})$ space. Due to lack of space, the algorithmic details are omitted. The purpose of redundancy removal is to eliminate the less trustworthy contained sequences in order to avoid possible false grouping later during the dense subgraph stage. Let S' be the set of non-redundant sequences, and let |S'| = n'.

 $^{^{3}}$ It is to be noted that the cutoffs mentioned as part of our approach throughout the paper are values that can be specified by the user as software parameters. The absolute values specified in the paper denote default parameter settings.

B. Detection of Connected Components

Definition 2: Two sequences are said to "overlap" if they share a local alignment such that the similarity is at least 30% and the alignment includes at least 80% of the longer sequence.

Problem 2: Given S', detect all maximal subsets (or "clusters") such that each sequence in a cluster overlaps with at least one other cluster member.

The PaCE parallel algorithm [19] was originally designed to address this problem although in the context of clustering DNA sequences. The key steps can be outlined as follows:

- 1) The algorithm uses the master-worker paradigm.
- 2) A distributed representation of a GST is generated using the input S' on all the worker processors such that each processor stores a unique $O(\frac{n^2 \ell}{p})$ portion of the tree.
- The master processor initializes the set of clusters such that each sequence is placed in a cluster of its own. The clustering is implemented using the union-find data structure [29] to allow for near-constant time *find()* and *merge()* operations.
- 4) An iterative process of data exchange between the master and workers is started. At each iteration, a worker processor is responsible for (i) sending new pairs of sequences that have a significantly long ($\geq \psi$) maximal match; these pairs are called *promising pairs* because they have a high likelihood of passing the overlap test, and (ii) computing alignments [23], [27] over the pairs of sequences that the master assigned to it. Alternatively, the master processor is responsible for (i) updating the current clustering based on the alignment results each worker returns, (ii) identifying maximal matching pairs that need alignment computation, and (iii) dynamically distributing the pending alignment workload to the worker processors.

Drastic savings in run-time are brought about by heuristic strategies. At any given time, if two sequences s_i and s_j are found to have an overlap, then their corresponding clusters are immediately merged. This is implemented using two find() and one union() operations. This transitive closure merging scheme is useful because later, if two sequences are reported as a promising pair and if the master processor locates them in the same cluster, then there is no necessity to compute their alignment. To maximize the chance that pairs leading to cluster merges are found earlier in the process, the dynamic promising pair detection algorithm generates them directly in the decreasing order of their maximal match length using an on-demand scheme as described in [19].

C. Bipartite Graph Generation

For each connected component generated in the previous phase, a bipartite graph consistent with the reductions described in Section III is to be generated. As long as the connected components are still small enough to fit in the memory of a single compute node, storing and analyzing a bipartite graph on a single node is a feasible option. For example, our implementation can handle a bipartite graph with up to a total of 16K vertices on a 512 MB RAM, or equivalently connected components with up to 8K vertices. For generating these bipartite graphs, however, all pairs of possible vertices need to be explored for a possible edge, although the maximal match heuristic can help reduce the work to some extent. For this reason, we parallelized bipartite graph generation using a modified version of the PaCE approach in which we apply only the maximal matching heuristic (and skip clustering). For the domain-based approach, the algorithm is much simpler as each fixed-length match identified is to be connected to their container sequences.

D. Dense Subgraph Detection in Bipartite Graphs

Once bipartite graphs consistent with the preferred reduction scheme are generated, the next step is to distribute them to multiple processors and apply the Shingle algorithm [12] on each bipartite graph serially. We made a few subtle modifications to the Shingle algorithm to suit the context. To assess the effectiveness of the algorithm's application it is necessary to understand the fundamentals of this algorithm. Like in the case of the PaCE algorithm, a thorough discussion is beyond the scope of this paper, and so we focus primarily on its major steps and the effect of its parameters on our dense subgraph problem. Most of the following discussions apply to both the duplicate (B_d) and match-based (B_m) bipartite approaches.

The Shingle Algorithm: Given a vertex v in an undirected bipartite graph $B = (V_l, V_r, E)$, $\Gamma(v)$ denotes the set of its out-links and is given by $\{u \mid (v, u) \in E\}$.

Definition 3: Given parameters (s, c), a "shingle" [7] of a vertex v is an arbitrary s-element subset of $\Gamma(v)$, and an "(s, c)-shingle set" of v is a set of c shingles of v.

Intuitively, two vertices sharing a shingle, by definition, share s of their out-links. In case of a dense bipartite subgraph, such pairs of vertices should be plenty on either side. The Shingle algorithm seeks to group such vertices together and use them for building dense subgraphs. Larger the value of s, lesser the probability that two vertices share a shingle, and vice versa. This implies that a smaller value of s is suited to enhance the chance of detecting not-so-dense subgraphs. The parameter c is intended to create the opposite effect. Also, it is not computationally practical to exhaustively compare all shingles of each pair of vertices, and the parameter coffers an alternative to restrict this computation space. This is achieved by using the min-wise independent permutation property [6]. Instead of generating c arbitrary shingles for a vertex v, the algorithm first generates c randomly sorted permutations of $\Gamma(v)$ and selects the s minimum elements from each permutation. Even if two vertices share a modest number of out-links, the randomness in this property will ensure that the probability of the vertices sharing a shingle is sufficiently high. This will be particularly helpful for detecting large subgraphs, as they are expected to be less dense. These



Fig. 4. Illustration of the two-pass Shingle algorithm.

parametric controls make the Shingle algorithm an ideal choice for our dense subgraph problem.

The algorithm is implemented in two-passes. See Figure 4 for an illustration.

- Pass I: An (s₁, c₁)-shingle set (denoted by S(v_i)) is generated for each vertex v_i ∈ V_l. For ease of implementation, each shingle is mapped to an integer using a hash function. The results are recorded as a 2-tuple <s(v_i), v_i>, where s(v_i) ∈ S(v_i). Let S₁ denote the union of all shingles generated in this pass. Next, vertices sharing the same shingle are grouped. This is achieved by sorting the tuples based on shingle values. The resulting tuple list is input to the second pass.
- **Pass II:** The algorithm reverses direction and generates an (s_2, c_2) -shingle set for each first level shingle $s(v_i)$. The result is a set of second level shingles S_2 , representing vertices from V_l .

In the final reporting step, all connected components (defined by S_2 shingle to S_1 shingle edges) are enumerated and their constituent vertices recorded. We implemented this using the union-find data structure. Two subsets of vertices A and Bare reported for each connected component, where A and Bdenote the sets of vertices from V_l and V_r respectively. For the global similarity-based graph approach, we output each $A \cup B$ as one dense subgraph provided it also satisfies the $\frac{|A \cap B|}{|A \cup B|} \ge \tau$ test. For the domain-based approach, we output B directly.

E. Implementation

We implemented our algorithms in C and MPI. The current version of our implementation only supports global similaritybased graph approach. To ensure code reuse wherever possible, we modularized the PaCE software code into separate modules for generalized suffix tree construction, maximal match detection, and the master-worker phase for clustering, and modified them for to work for ORF/amino acid sequences. The modularized codes were used as the implementation for the redundancy removal and connected component detection phases. For bipartite generation we used the maximal match detection code while implementing a new code for creating the adjacency list corresponding to each bipartite graph. For the dense subgraph detection using the Shingle algorithm, we implemented the code from scratch. Additional scripts were written to streamline the individual phases into an automated pipeline.

V. EXPERIMENTAL RESULTS

Our experimental studies were conducted on two platforms: i) a 512-node BlueGene/L supercomputer with each node containing two 700 MHz PPC cores and 512 MB RAM; and ii) a 24-node Linux commodity cluster with a gigabit ethernet interconnect and each node containing 8 2.33GHz Xeon CPUs and an 8 GB RAM. All our experiments on the BlueGene/L were run using the co-processor mode. For the redundancy removal (RR) and connected component detection (CCD) phases, we use the BlueGene/L supercomputer. For the dense subgraph detection (DSD) phase, we used the Linux cluster. The rationale for using two different platforms is multi-fold. The BlueGene/L platform enabled us to conduct scalability studies up to 512 processors; while the Linux cluster provided a higher per node memory that was required by the DSD code on connected components with more than 8K vertices. Also, the 64-bit CPUs in the Linux cluster were more suited for our implementation of the hash function and random number generation functionalities.

Data Preparation: The data for our experiments were downloaded from the CAMERA web portal (http://camera. calit2.net/), which contains a total of 28.6 million ORFs spanning different data classes: NCBI-nr [32], PG [32], TGI-EST [24], ENS [4], [5] and environmental sequences from the GOS project. The database also hosts the collection of predicted ORF families (henceforth, referred to as "clusters") reported by their team [33]. We extracted two sets of data from an arbitrary set of clusters such that all sequences from the selected clusters were used in the analysis. The first data set contained 160.000 ORFs spanning 221 clusters with an average sequence length of 163 amino acid residues. Subsets of this data (10K, 20K, 40K, 80K) were used for performance evaluation. The second data set contained 22,186 ORFs spanning one large cluster with an average sequence length of 256 residues.

Qualitative Evaluation: We conducted quality evaluation on the 22K and 160K data sets. The RR step reduced the two sets to 21K and 138K respectively. All subsequent analysis were conducted only on these non-redundant sets. A dense subgraph minimum size cutoff of 5 was used in our method, and a fine tuned set of parameters of (5,300) was used for (s, c). The CCD code on the 138K data produced 1.8K connected components of size 5 or more. These components included 95K (out of the 138K) sequences. Each component

 TABLE I

 Table summary of our qualitative assessment on the 22K and 160K data sets. Results are reported for only those connected components containing 5 sequences or more.

#Input seq.	#NR ^a seq.	#CC ^b	#DS ^c	#Seq in DS	Mean degree	Mean density	Size of largest DS
160,000	138,633	1,861	850	66,083	26	76%	13,263
22,186	21,348	1	134	11,524	20	78%	6,828

a "NR" stands for non-redundant.

^b "CC" stands for connected components.

^c "DS" stands for dense subgraphs.



Fig. 5. Distribution of our dense subgraphs as a function of their sizes. The largest dense subgraph contains over 7K sequences and is not shown in the plot.

was then individually input to the DSD step. Overall, 850 dense subgraphs (DS) were detected, which covered 66K sequences, implying on an average \sim 78 sequences per DS. Each sequence (or vertex) on average was connected to 26 other sequences. We also calculated the observed "density" of each DS. For a DS with *m* nodes, density is given by $\frac{\#\text{mean degree}}{m-1}$. The mean density over all the reported dense subgraphs was observed to be 76%, implying high connectivity among the nodes. This demonstrates the ability of our method to detect high quality dense subgraphs regardless of their individual sizes. Similar study was carried out for the 22K data set. The results are subgraphs as a function of their sizes for the 22K data. The distribution is skewed as expected. The largest cluster had over 7K sequences (not shown in the plot).

For comparison we used the results from the GOS project. The GOS clustering involves several stages. After the initial clustering (called core set identification), the similarity criteria are relaxed to accommodate far related proteins, and is followed by a model-based clustering, which uses a training set to allow for incorporation of knowledge on known protein families. Since we perform only a sequence similarity based clustering a direct comparison with the final GOS clustering cannot be fair. Nevertheless, it is only the sensitivity that is affected by this discrepancy in the two methodologies. To validate this, we performed the following assessment.

For the 160K data set, the GOS clustering had 221 final clusters, while our approach detected 850 DS. For 22K one

GOS cluster was fragmented into 134 DS. This is expected as per the above sensitivity claim. To evaluate agreements and discrepancies, we treated our DS-based clustering as the "Test" clustering and the GOS clustering as the "Benchmark". A pair of sequences is called (i) "True Positive" (TP) if they are clustered together in both schemes; (ii) "True Negative" (TN) if they separated in both schemes; (iii) "False Positive" (FP) if they are clustered together in Test but not in Benchmark; and (iv) "False Negative" (FN) if they are clustered together in Benchmark but not in Test.

Precision Rate (PR) =
$$\frac{\text{TP}}{\text{TP+FP}}$$
 (1)

Sensitivity (SE) =
$$\frac{TP}{TP+FN}$$
 (2)

Overlap Quality (OQ) =
$$\frac{\text{TP}}{\text{TP}+\text{FP}+\text{FN}}$$
 (3)

Correlation Coefficient (CC) =

$$\frac{\text{TP.TN-FP.FN}}{\sqrt{(\text{TP+FP}).(\text{TN+FN}).(\text{TP+FN}).(\text{TN+FP})}}$$
(4)

Ideally, OQ=PR=SE=CC=100%. For a valid comparison, we calculated the above measures by observing the distribution of sequences that were included in the clustering under both schemes.

For 160K the results of comparison are as follows:

PR=95.75%; SE=56.89%; OQ=55.49%; CC=73.04%

While SE is low as expected, the high value of PR suggests that most of our clustering is also preserved in the Benchmark. The high prediction accuracy of our method along with its effectiveness in detecting high density subgraphs is suited for generating a more accurate core set. We believe our dense subgraph detection scheme followed subsequently by the set of thorough evaluations performed in the GOS project can provide an ideal combination for the protein family identification problem.

Performance Evaluation: For performance evaluation we used the 160K data set and its subsets. The RR and CCD implementations were tested on 512 BG/L nodes (in coprocessor mode). The DSD code was tested on the 24-node Linux commodity cluster.

Figure 6a shows the combined run-time for the RR and CCD phases as a function of processor size. The 160K input run on 512 processors completed in 3h 20m. Between the two



Fig. 6. Parallel run-times (in seconds) on BG/L for the RR and CCD phases as a function of (a) processor size, and (b) input size.



Fig. 7. (a) Speedup of the RR and CCD phases for varying input sizes on 512 nodes of a BlueGene/L supercomputer. The minimum processor size used was 32 nodes, and hence the speedup figures are calculated relative to a 32-node system. Also, due to a technical (resource management) restriction it was not possible to launch jobs on a 256 node system. (b) Serial run-time for dense subgraph detection as a function of input size and (s, c) parameter values.

phases, the RR phase accounted for more than 90% of all run-times. This is as expected because detection of redundant sequences do not lead to any clustering. The CCD phase is faster when compared to the RR phase because successfully tested alignments lead to merging of clusters, and thereby result in drastic reductions in alignment work. For example, on the 40K input 168 million promising pairs were generated based on maximal matches of length 10 residues, and of which only 7 million pairs were selected for alignment computations. This corresponds to a 99% in work reduction when compared to any scheme that deploys an all-versus-approach approach $\binom{40K}{2} \approx 800$ million alignment computations). Figure 6b shows run-times as a function of the input size. While the run-time has an asymptotic worst-case quadratic complexity, the efficacy of the clustering heuristic technique varies with the input data.

Figure 7a shows the speedup of executing the RR and CCD phases. As can be observed, the speedup figures are closer to linear for larger input sizes; while for higher number of processors (e.g., 128 to 512) there is only a modest increase

in speedup (e.g., from 3.6 to 6.7 vs. an ideal 4 to 16). Upon investigation, we found that this loss in expected speedup was due to the CCD phase. As a concrete example, consider the 80K input case, for which the respective run-times for the RR and CCD are tabulated in Table II. As can be noted, the scaling of the RR phase, which is the most dominant phase, is mostly linear; whereas, there is poor scaling of the CCD phase. During the CCD phase, the clustering heuristic eliminates an overwhelming majority (more than 99.9%) of the promising pairs generated based on maximal matches. As a result, the work generated by the worker processors were being too aggressively filtered out on the master node, thereby leaving very little alignment work to be redistributed to the worker nodes. This is both a beneficial and undesirable outcome - beneficial because only an insignificant fraction of the overall pairs generated are actually aligned thereby drastically reducing the overall time to solution; and undesirable because the work reduction adversely affects the scaling. A more aggressive work generation scheme is required to compensate for work loss.

TABLE II RUN-TIMES (IN SECONDS) FOR THE RR AND CCD PHASES FOR THE 80K INPUT CASE.

Phase	Number of processors						
	32	64	128	512			
RR CCD	17,476 1,068	10,296 777	4,560 528	2,207 670			

The dense subgraph detection phase was relatively faster when compared to the previous phases. Even on the largest connected component recorded in our experiments (\sim 20K), the serial code took less than 10 minutes on a single processor of the linux cluster. The average run-time for connected components generated for the 160K input was \sim 3 minutes. Because of the short run-times for each connected component, we grouped multiple connected components into batches of roughly the same size and distributed the batches across processors. The plot in Figure 7b shows the run-time statistics on these groupings as a function of input size and shingle parameters, s and c. It can be observed that the run-time increases with increasing values of the c parameter. This is because more shingles will be generated and hence increase the overall workload.

VI. FUTURE WORK

We plan to continue the development effort in several directions.

- **Quality testing:** The effect of similarity cutoffs and other parameters on the quality of the protein family prediction is to be studied. More extensive qualitative validations are necessary to ascertain the merits of our approach for large-scale identification of protein families in metagenomic data sets. In addition, we plan to implement, test and compare the domain-based family detection approach proposed in this paper.
- **Parallelization of the Shingle algorithm:** While time is not likely to be an issue for the dense subgraph detection phase (at least for the range of inputs tested), our goal is to parallelize the shingle step to address the need for memory. Currently, if all shingles on either side of the bipartite graph are unique then the peak space requirement is proportional to $O(m \times c^2)$, where *m* is the cardinality of the smaller of the partitions in the bipartite graph.
- Large-scale application: While our experiments on small to medium-sized inputs show promising performance and quality results, tens of millions of new sequence data are becoming rapidly available. Perhaps a greater challenge will be to extend the methodologies developed here to solve problems of much larger scales with improved quality.

VII. CONCLUSIONS

We presented a novel parallel approach for protein family identification. This is a challenging problem of growing significance, and is an ideal application to benefit from supercomputing technologies. We draw from the strengths of two methods (PaCE and Shingle) previously developed for other applications, and apply them in context of a new problem. Overall the results presented in this paper demonstrate the high potential and positive impact of parallelism for metagenomic protein family identification. They have also pointed us to many new research directions for further investigation.

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